

The Burden of Retinal Degenerative Diseases

WHITEPAPER SERIES IN OPHTHALMOLOGY

2023



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Retinal degenerative diseases, like diabetic retinopathy, glaucoma, and age-related macular degeneration (AMD), are leading causes of vision loss and blindness worldwide¹.

The Burden of Retinal Degenerative Diseases

Vision impairment and blindness have a substantial impact on people's ability to perform everyday tasks and can affect their quality of life. In fact, vision impairment reduces mobility, affects mental wellbeing, exacerbates the risk of dementia, increases the likelihood of falls and road traffic crashes, increases the need for social care, and ultimately leads to higher mortality rates².

Globally, at least 2.2 billion people have a vision impairment or blindness, of whom at least 1 billion have a vision impairment that could have been prevented or has yet to be addressed². Moreover, in the "World report on the vision" from World Health Organization (WHO), it was reported that 11.9 million people globally have a moderate or severe vision impairment or blindness due to glaucoma, diabetic retinopathy, and trachoma that could have been prevented³.

The estimated costs of preventing the vision impairment in these 11.9 million would have been US\$5.8 billion³. Despite the scarce economic estimates, it was suggested that the annual global productivity loss from vision impairment is approximately US\$410.7 billion purchasing power parity⁴. That way the prevention of vision impairment represents a significant opportunity to reduce the substantial personal and societal burden associated with vision impairment and blindness.

The pathophysiology of diabetic retinopathy, glaucoma, and AMD is still not completely understood, and novel biomarkers for diagnosis and disease progression are needed. Early diagnosis and intervention on patients with retinal degenerative disease would be key to improve the outcomes of treatment. Despite the increasing interest of the scientific community in trying to elucidate the mechanisms underlying retinal dysfunction and degeneration, that knowledge is yet to translate into effective therapies for these retinal degenerative diseases. That way, there is a clear unmet clinical need for new and more effective therapeutic strategies⁵.

Diabetic Retinopathy

Diabetic retinopathy is a significant complication of diabetes mellitus and a leading cause of vision impairment and blindness in working-age adults⁶.

Globally, 3.28 millions of people present visual impairment and 1.07 million are blind due to diabetic retinopathy ⁴ (4). A major hallmark of the disease is the blood-retinal barrier breakdown. The current available Therapeutic strategies mainly target neovascularization through the use of anti-vascular endothelial growth factor (VEGF) therapies, laser treatment, and surgery⁷. Nevertheless, diabetic retinopathy is now considered a neurovascular disease in which a low-grade chronic inflammatory environment contributes to blood-retinal barrier breakdown and retinal neural dysfunction^{8,9}. In fact, the inducible nitric oxide synthase isoform, which is highly involved in inflammation processes, is a key mediator of blood-retinal barrier breakdown triggered by diabetes¹⁰.

Glaucoma

Glaucoma is a leading cause of irreversible blindness and is characterized by optic nerve damage and retinal ganglion cell death¹¹. It was calculated that glaucoma led to visual impairment and blindness of 4.13 and 3.61 million people in 2020, respectively⁴. Elevated intraocular pressure (IOP) is an important risk factor in glaucoma and the only one that is modifiable¹². Indeed, current treatments are directed towards IOP lowering. However, many patients continue to lose vision despite successful IOP control¹³. Therefore, new and more effective treatments are an emergent clinical need, and retinal neuroprotection has been considered to be an additional therapy¹⁴. Moreover, it has been described that microglia-mediated neuroinflammation further contributes to retinal neurodegeneration and the underlying mechanisms have been studied to devise novel neuroprotective strategies for glaucoma¹⁵. That way a new therapeutic approach to glaucoma should be not only focused on IOP lowering but also on protecting retinal ganglion cell from glaucomatous damage^{16,17}. The control of microglia-mediated neuroinflammation was demonstrated to confer neuroprotection to the retina^{18,19}.

Age-related macular degeneration (AMD)

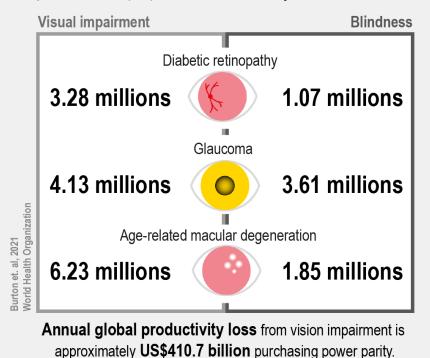
AMD is a major cause of vision loss worldwide and the leading cause of blindness in the elderly²⁰. It is responsible for visual impairment in 6.23 million people and blindness in 1.85 million people⁴. AMD is classified in two forms²¹:

- "Early/dry" form: characterized by the accumulation of cellular debris in the subretinal space, beneath the retinal pigment epithelium (RPE), called drusens. The disease can then progress to RPE and photoreceptor loss;
- "Advanced dry/wet" form: characterized by the presence of geographic atrophy (dry) due to RPE and photoreceptor cell death and/or choroidal neovascularization (wet), with new blood vessels arising from the choroid through the RPE layer into the outer retina, leading to photoreceptor dysfunction and degeneration

The dry form of AMD and geographic atrophy are currently untreatable, while VEGF inhibition therapy is the best available treatment for wet AMD²². The anti-VEGF therapies present a significant burden for the patients due to the need for repeated intravitreal injections²³. That way alternative treatment strategies have been proposed in order to ease the treatment burden by reducing the number of injections needed ^{24,25}.

In Vivo Models

The development of several animal models of disease mimicking some features present in retinal degenerative diseases have been significantly contributing to unraveling



Globally, **2.2 billion** people have a vision impairment or blindness.

FIGURE 1:

Global estimates of number of people affected by retinal degenerative diseases, namely diabetic retinopathy, glaucoma and age-related macular degeneration, that cause vision impairment and blindness.

important aspects in the understanding of the pathogenesis of the retinal

disease. The development of in vivo models also allows testing new and more efficacious possible therapeutic strategies for retinal degenerative diseases²⁶. In diabetic retinopathy research, there are several animal models that rely on the induction of both type I diabetes and type 2 diabetes. One of the commonly used animal models of diabetic retinopathy is the streptozotocin (STZ)-induced type I diabetes. This model has been used to give insight into the action mechanisms of new and already used treatments in the clinics ^{27,28}. Goto-Kakizaki (GK) animals are a type 2 diabetes animal model characterized by early and relatively stable mild hyperglycemia, hyperinsulinemia, and insulin resistance ²⁹. This model mimics some features present in diabetic retinopathy as increased nitric oxide production, early inner blood-retinal barrier breakdown and migration of activated microglial cells from the retina to the choroid ^{30,31}. Regarding glaucoma, since elevated IOP is the main risk factor, relevant animal models for glaucoma have been developed around the challenge of producing experimental IOP elevation. However, other non-related IOP animal models have been developed³². Glaucoma animal models lead to retinal ganglion cells (RGCs) dysfunction and loss, and optic nerve axonal transport impairment, mimicking some features of the disease. Ocular hypertension (OHT) model, based on laser photocoagulation of the trabecular meshwork and perilimbal and episcleral veins³³, episcleral vein cauterization model³⁴, and microbeads injection in the anterior chamber³⁵ are animal models based on the blockage of aqueous humor drainage, increasing IOP, and ultimately leading to RGC loss and damage of the optic nerve. Moreover, pressure-independent animal models, like optic nerve crush, optic nerve transection, and retinal ischemiareperfusion (I-R) injury³², have been used to model normal-tension glaucoma, and they have provided insights into the neurodegenerative mechanisms of RGC loss.

The animal models that mimic some features of AMD are focused on both forms of the disease, "dry" and "wet". Complement factor pathway genetic models (e.g. Ccl2-/- and Ccr2 mice)³⁶ and light-induced retinal degeneration model³⁷ have being used to study the advanced dry geographic form of AMD, inducing photoreceptor death, subretinal drusen-like accumulations, thickening of Bruch's membrane and immune activation. The laser-induced choroidal neovascularization animal model is the most used model for "wet" AMD, which induces Bruch's membrane rupture, inducing the growth of new choroidal vessels into the subretinal area³⁸.

Monitoring of Retinal Degenerative Diseases

Accurate monitoring of retinal degenerative diseases in patients is vital to preserve their visual function.

Likewise, disease progression can be assessed using similar but adapted methods in animal models.

Monitoring of Retinal Degenerative Diseases

- The leakage of the blood-retinal barrier can be assessed in vivo by fluorescein angiography, which is widely used to assess alterations in the retinal vasculature in diabetic retinopathy and age-related macular degeneration patients³⁹.
- Retinal function and structure can be easily assessed in vivo in animal models of disease by electroretinography (ERG) and optical coherence tomography (OCT),

respectively. Significant progress have been achieved in retinal imaging techniques that have enabled clinicians to detect structural changes in patients⁴⁰. OCT is a non-invasive procedure used to visualize the anterior and posterior segments of the eye at high resolution that likewise is used in animals^{41,42}.

 Assessment of visual function, used in the clinic has also been adapted and can be performed in animal models using quantitative optomotor response ⁴³.

Optical Coherence Tomography (OCT)

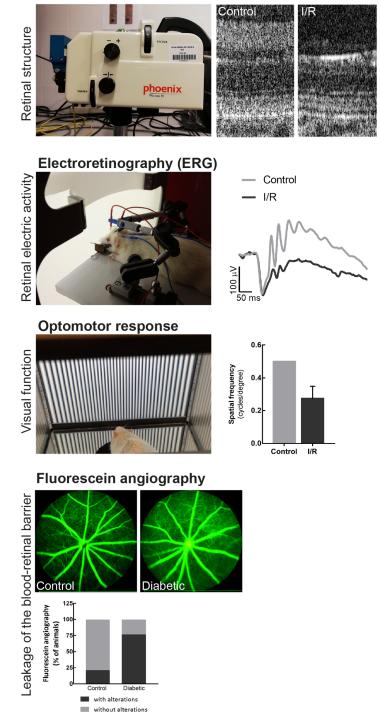


FIGURE 2:

Disease progression in animal models of disease can be performed by optical coherence tomography (OCT, to assess retinal structure), electroretinography (ERG, to assess retinal function), and quantitative optomotor response (OMR, to assess visual function) and fluorescein angiography (to assess leakage of the blood-retinal barrier).

The retina as a Window to the Brain

The concept of "the retina as a window to the brain" arose several years ago since eye examination, and namely the retina, with non-invasive approaches has been considered an additional tool for the early diagnosis of neurodegenerative diseases⁴⁴, such as Alzheimer's disease^{45,46} and Parkinson's disease⁴⁷. In fact, several studies in Alzheimer's disease animal models48 and patients49.50 demonstrated a thinning of the retina assessed by OCT. These studies suggest that the assessment of retinal thickness by OCT may be useful for an early diagnosis of Alzheimer's disease. Similar findings have been reported in Parkinson's disease patients^{51,52}.

References

1. Bourne RRA, Flaxman SR, Braithwaite T, Cicinelli MV, Das A, Jonas JB, et al. Magnitude, temporal trends, and projections of the global prevalence of blindness and distance and near vision impairment: a systematic review and meta-analysis. The Lancet Global health. 2017;5(9):e888-e97.

2. [WHO] WHO. Blindness and Vision Impairment 2021. Available from: https://www.who.int/news-room/fact-sheets/detail/blindness-and-visualimpairment.

3. [WHO] WHO. Blindness and Vision Impairment - World report on vision 2019. Available from: https://www.who.int/publications/i/item/world-report-on-vision.

4. Burton MJ, Ramke J, Marques AP, Bourne RRA, Congdon N, Jones I, et al. The Lancet Global Health Commission on Global Eye Health: vision beyond 2020. The Lancet Global health. 2021.

 Cursiefen C, Cordeiro F, Cunha-Vaz J, Wheeler-Schilling T, Scholl Hendrik PN. Unmet Needs in Ophthalmology: A European Vision Institute-Consensus Roadmap 2019–2025. Ophthalmic Research. 2019;62(3):123-33.
 Santiago AR, Boia R, Aires ID, Ambrosio AF, Fernandes R. Sweet Stress: Coping With Vascular Dysfunction in Diabetic Retinopathy. Frontiers in physiology. 2018;9:820.

7. Marozas LM, Fort PE. Diabetic Retinopathy-Update on Prevention Techniques, Present Therapies, and New Leads. US ophthalmic review. 2014;7(1):54-8.

 Villarroel M, Ciudin A, Hernandez C, Simo R. Neurodegeneration: An early event of diabetic retinopathy. World journal of diabetes. 2010;1(2):57-64.

 Rubsam A, Parikh S, Fort PE. Role of Inflammation in Diabetic Retinopathy. International journal of molecular sciences. 2018;19(4).
 Leal EC, Manivannan A, Hosoya K, Terasaki T, Cunha-Vaz J, Ambrosio AF, et al. Inducible nitric oxide synthase isoform is a key mediator of leukostasis and blood-retinal barrier breakdown in diabetic retinopathy. Investigative ophthalmology & visual science. 2007;48(11):5257-65.

11. Weinreb RN, Aung T, Medeiros FA. The pathophysiology and treatment of glaucoma: a review. Jama. 2014;311(18):1901-11.

12. Coleman AL, Kodjebacheva G. Risk factors for glaucoma needing more attention. The open ophthalmology journal. 2009;3:38-42.

 Leske MC, Heijl A, Hussein M, Bengtsson B, Hyman L, Komaroff E, et al. Factors for glaucoma progression and the effect of treatment: the early manifest glaucoma trial. Archives of ophthalmology. 2003;121(1):48-56.
 Boia R, Ruzafa N, Aires ID, Pereiro X, Ambrosio AF, Vecino E, et al. Neuroprotective Strategies for Retinal Ganglion Cell Degeneration: Current Status and Challenges Ahead. International journal of molecular sciences.

 Madeira MH, Boia R, Santos PF, Ambrosio AF, Santiago AR.
 Contribution of microglia-mediated neuroinflammation to retinal degenerative diseases. Mediators of inflammation. 2015;2015:673090.
 Galvao J, Elvas F, Martins T, Cordeiro MF, Ambrosio AF, Santiago AR.
 Adenosine A3 receptor activation is neuroprotective against retinal neurodegeneration. Experimental eye research. 2015;140:65-74.

2020:21(7).

 Boia R, Salinas-Navarro M, Gallego-Ortega A, Galindo-Romero C, Aires ID, Agudo-Barriuso M, et al. Activation of adenosine A3 receptor protects retinal ganglion cells from degeneration induced by ocular hypertension. Cell death & disease. 2020;11(5):401.

18. Madeira MH, Boia R, Elvas F, Martins T, Cunha RA, Ambrosio AF, et al. Selective A2A receptor antagonist prevents microglia-mediated neuroinflammation and protects retinal ganglion cells from high intraocular pressure-induced transient ischemic injury. Translational research : the journal of laboratory and clinical medicine. 2016;169:112-28.

19. Boia R, Elvas F, Madeira MH, Aires ID, Rodrigues-Neves AC, Tralhao P, et al. Treatment with A2A receptor antagonist KW6002 and caffeine intake regulate microglia reactivity and protect retina against transient ischemic damage. Cell death & disease. 2017;8(10):e3065.

20. Wong WL, Su X, Li X, Cheung CM, Klein R, Cheng CY, et al. Global prevalence of age-related macular degeneration and disease burden projection for 2020 and 2040: a systematic review and meta-analysis. The Lancet Global health. 2014;2(2):e106-16.

21. de Jong EK, Geerlings MJ, den Hollander AI. Age-related macular degeneration. 2020:155-80.

22. Moutray T, Chakravarthy U. Age-related macular degeneration: current treatment and future options. Therapeutic advances in chronic disease. 2011;2(5):325-31.

23. Edelhauser HF, Rowe-Rendleman CL, Robinson MR, Dawson DG, Chader GJ, Grossniklaus HE, et al. Ophthalmic drug delivery systems for the treatment of retinal diseases: basic research to clinical applications. Investigative ophthalmology & visual science. 2010;51(11):5403-20.

24. Campochiaro PA, Marcus DM, Awh CC, Regillo C, Adamis AP, Bantseev V, et al. The Port Delivery System with Ranibizumab for Neovascular Age-Related Macular Degeneration. Ophthalmology. 2019;126(8):1141-54.

25. Regillo C, editor Long-acting drug delivery in nAMD: ARCHWAY phase 3 results. EURETINA 2020 virtual; 2020.

26. Fletcher EL, Jobling Al, Vessey KA, Luu C, Guymer RH, Baird PN. Animal models of retinal disease. Progress in molecular biology and translational science. 2011;100:211-86.

27. Goncalves A, Marques C, Leal E, Ribeiro CF, Reis F, Ambrosio AF, et al. Dipeptidyl peptidase-IV inhibition prevents blood-retinal barrier breakdown, inflammation and neuronal cell death in the retina of type 1 diabetic rats. Biochimica et biophysica acta. 2014;1842(9):1454-63.

28. Aires ID, Madeira MH, Boia R, Rodrigues-Neves AC, Martins JM, Ambrosio AF, et al. Intravitreal injection of adenosine A2A receptor antagonist reduces neuroinflammation, vascular leakage and cell death in the retina of diabetic mice. Scientific reports. 2019;9(1):17207.

29. King AJ. The use of animal models in diabetes research. British journal of pharmacology. 2012;166(3):877-94.

30. Carmo A, Cunha-Vaz JG, Carvalho AP, Lopes MC. Nitric oxide synthase activity in retinas from non-insulin-dependent diabetic Goto-Kakizaki rats: correlation with blood-retinal barrier permeability. Nitric oxide : biology and chemistry. 2000;4(6):590-6.

31. Campos A, Martins J, Campos EJ, Silva R, Ambrosio AF. Choroidal and retinal structural, cellular and vascular changes in a rat model of Type 2 diabetes. Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie. 2020;132:110811.

32. Johnson TV, Tomarev SI. Rodent models of glaucoma. Brain research bulletin. 2010;81(2-3):349-58.

33. Salinas-Navarro M, Alarcon-Martinez L, Valiente-Soriano FJ, Jimenez-Lopez M, Mayor-Torroglosa S, Aviles-Trigueros M, et al. Ocular hypertension impairs optic nerve axonal transport leading to progressive retinal ganglion cell degeneration. Experimental eye research. 2010;90(1):168-83.

34. Urcola JH, Hernandez M, Vecino E. Three experimental glaucoma models in rats: comparison of the effects of intraocular pressure elevation on retinal ganglion cell size and death. Experimental eye research. 2006;83(2):429-37.

35. Morgan JE, Tribble JR. Microbead models in glaucoma. Experimental eye research. 2015;141:9-14.

36. Ambati J, Anand A, Fernandez S, Sakurai E, Lynn BC, Kuziel WA, et al. An animal model of age-related macular degeneration in senescent Ccl-2or Ccr-2-deficient mice. Nature medicine. 2003;9(11):1390-7.

 Scholz R, Sobotka M, Caramoy A, Stempfl T, Moehle C, Langmann T. Minocycline counter-regulates pro-inflammatory microglia responses in the retina and protects from degeneration. Journal of neuroinflammation. 2015;12:209.

 Gong Y, Li J, Sun Y, Fu Z, Liu CH, Evans L, et al. Optimization of an Image-Guided Laser-Induced Choroidal Neovascularization Model in Mice. PloS one. 2015;10(7):e0132643. 29. King AJ. The use of animal models in diabetes research. British journal of pharmacology. 2012;166(3):877-94.

30. Carmo A, Cunha-Vaz JG, Carvalho AP, Lopes MC. Nitric oxide synthase activity in retinas from non-insulin-dependent diabetic Goto-Kakizaki rats: correlation with blood-retinal barrier permeability. Nitric oxide : biology and chemistry. 2000;4(6):590-6.

31. Campos A, Martins J, Campos EJ, Silva R, Ambrosio AF. Choroidal and retinal structural, cellular and vascular changes in a rat model of Type 2 diabetes. Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie. 2020;132:110811.

32. Johnson TV, Tomarev SI. Rodent models of glaucoma. Brain research bulletin. 2010;81(2-3):349-58.

33. Salinas-Navarro M, Alarcon-Martinez L, Valiente-Soriano FJ, Jimenez-Lopez M, Mayor-Torroglosa S, Aviles-Trigueros M, et al. Ocular hypertension impairs optic nerve axonal transport leading to progressive retinal ganglion cell degeneration. Experimental eye research. 2010;90(1):168-83.

34. Urcola JH, Hernandez M, Vecino E. Three experimental glaucoma models in rats: comparison of the effects of intraocular pressure elevation on retinal ganglion cell size and death. Experimental eye research. 2006;83(2):429-37.

35. Morgan JE, Tribble JR. Microbead models in glaucoma. Experimental eye research. 2015;141:9-14.

36. Ambati J, Anand A, Fernandez S, Sakurai E, Lynn BC, Kuziel WA, et al. An animal model of age-related macular degeneration in senescent Ccl-2or Ccr-2-deficient mice. Nature medicine. 2003;9(11):1390-7.

37. Scholz R, Sobotka M, Caramoy A, Stempfl T, Moehle C, Langmann T. Minocycline counter-regulates pro-inflammatory microglia responses in the retina and protects from degeneration. Journal of neuroinflammation. 2015;12:209.

 Gong Y, Li J, Sun Y, Fu Z, Liu CH, Evans L, et al. Optimization of an Image-Guided Laser-Induced Choroidal Neovascularization Model in Mice. PloS one. 2015;10(7):e0132643.

39. Chang B. Mouse models for studies of retinal degeneration and diseases. Methods in molecular biology. 2013;935:27-39.

40. Li Y, Xia X, Paulus YM. Advances in Retinal Optical Imaging. Photonics. 2018;5(2).

41. Kawaguchi I, Higashide T, Ohkubo S, Takeda H, Sugiyama K. In vivo imaging and quantitative evaluation of the rat retinal nerve fiber layer using scanning laser ophthalmoscopy. Investigative ophthalmology & visual science. 2006;47(7):2911-6.

42. Nagata A, Higashide T, Ohkubo S, Takeda H, Sugiyama K. In Vivo Quantitative Evaluation of the Rat Retinal Nerve Fiber Layer with Optical Coherence Tomography. Investigative Opthalmology & Visual Science. 2009;50(6):2809.

43. Abdeljalil J, Hamid M, Abdel-mouttalib O, Stéphane R, Raymond R, Johan A, et al. The optomotor response: A robust first-line visual screening method for mice. Vision research. 2005;45(11):1439-46.

44. London A, Benhar I, Schwartz M. The retina as a window to the brain—
from eye research to CNS disorders. Nature Reviews Neurology.
2012;9(1):44-53.

45. Colligris P, Perez de Lara MJ, Colligris B, Pintor J. Ocular Manifestations of Alzheimer's and Other Neurodegenerative Diseases: The Prospect of the Eye as a Tool for the Early Diagnosis of Alzheimer's Disease. Journal of Ophthalmology. 2018;2018:1-12. 46. Chiquita S, Rodrigues-Neves AC, Baptista FI, Carecho R, Moreira PI, Castelo-Branco M, et al. The Retina as a Window or Mirror of the Brain Changes Detected in Alzheimer's Disease: Critical Aspects to Unravel. Molecular Neurobiology. 2019;56(8):5416-35.

47. Moreno-Ramos T, Benito-León J, Villarejo A, Bermejo-Pareja F. Retinal Nerve Fiber Layer Thinning in Dementia Associated with Parkinson's Disease, Dementia with Lewy Bodies, and Alzheimer's Disease. Journal of Alzheimer's Disease. 2013;34(3):659-64.

48. Chiquita S, Campos EJ, Castelhano J, Ribeiro M, Sereno J, Moreira PI, et al. Retinal thinning of inner sub-layers is associated with cortical atrophy in a mouse model of Alzheimer's disease: a longitudinal multimodal in vivo study. Alzheimer's research & therapy. 2019;11(1):90.

49. Paquet C, Boissonnot M, Roger F, Dighiero P, Gil R, Hugon J. Abnormal retinal thickness in patients with mild cognitive impairment and Alzheimer's disease. Neuroscience letters. 2007;420(2):97-9.

50. Kesler A, Vakhapova V, Korczyn AD, Naftaliev E, Neudorfer M. Retinal thickness in patients with mild cognitive impairment and Alzheimer's disease. Clinical neurology and neurosurgery. 2011;113(7):523-6.

51. Bayhan HA, Aslan Bayhan S, Tanik N, Gurdal C. The association of spectral-domain optical coherence tomography determined ganglion cell complex parameters and disease severity in Parkinson's disease. Current eye research. 2014;39(11):1117-22.

52. Garcia-Martin E, Larrosa JM, Polo V, Satue M, Marques ML, Alarcia R, et al. Distribution of retinal layer atrophy in patients with Parkinson disease and association with disease severity and duration. American journal of ophthalmology. 2014;157(2):470-8 e2.

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